## Smartphone-Integrated Point-of-Care Sensor for Rapid and Sensitive Malaria Detection in Resource-Limited Settings

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## **Supplementary Information**

Figure S1. Primer Temperature Screen. Each set of primers was tested with four different thermocycling conditions. Primers were reacted in 50 mM KCl, 5 mM ammonium sulfate, 5% DMSO and 0.1% tween20 for 20 cycles with 30 seconds at the high (denaturing temperature) followed by 60 seconds at the low (annealing temperature).

"Click" suitable alkyne	Chem. Structure	$2^{nd}$ order rate constant (M <sup>-1</sup> s <sup>-1</sup> )
Dibenzocyclooctyne (DBCO)		0.31
Dibenzocyclooctyne (DIBO)	Q <sub>R</sub>	0.057
Cyclooctyne (OCT)	C C R	0.0024

**Figure S2. SPAAC-Ligation Reaction Groups.** Dibenzocyclooctyne (DBCO/DIBO) and Cyclooctyne (OCT) reactive groups react with azide to form ligated nucleic acid. Groups are listed in order from fastest to slowest reaction rate.



**Figure S3. Comparison of SPAAC Reaction Groups.** 27/31nts primer sets labeled with either DBCO, DIBO, or OCT reactive groups were tested in samples containing specific concentrations of target DNA and 25% bovine serum, 50 mM KCl, 20 mM ammonium sulfate, 5% DMSO, and 0.1% tween-20. Samples were incubated at 95°C for 10 minutes followed by 50 cycles of denaturing temperature (66°C) for 30 seconds followed by annealing temperature (31°C) for 60 seconds (Standard Thermocycling Time (STT), left) or denaturing for 20 seconds followed by annealing for 45 seconds (Reduced Thermocycling Time (RTT), right). All samples were tested in duplicate.

Target (pM)	GC/pL	-μA/s	App Result	App Concentration	<b>Relative Error</b>
1500	841	0.407	Negative	BLoQ	NA
700	392	0.447	Negative	BLoQ	NA
100	56.0	0.318	Negative	BLoQ	NA
25	14.0	0.215	Positive	21.7 GC/pL	-55.0%
5	2.80	0.0559	Positive	0.8 GC/pL	71.4%
1	0.560	0.0172	Positive	0.5 GC/pL	10.7%
0	0	0.0588	Positive	ALoQ	NA

Table S1. Comparison of Chronoamperometric Results Generated by PSTrace v.5.8 or the Giner Smartphone Application.

Samples were spiked with target at concentrations ranging from 1500 pM to 1 pM with unspiked sample serving as a negative control. Each sample was first measured using a potentiostat connected to a laptop running PSTrace v.5.8 software (PalmSens). Samples were then tested using the Giner smartphone application to determine the accuracy of the reported values. Only 3/6 positive samples returned calculated sample concentrations. Highly positive samples were reported as negative while the negative sample was reported as positive.